Cytoadhesion of *Plasmodium falciparum*-infected erythrocytes to chondroitin-4-sulfate is cooperative and shear enhanced

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**Key Points**

- Cytoadhesion of parasitized erythrocytes depends on the intermolecular distance between neighboring chondroitin-4-sulfate molecules.
- VAR2CSA is an allosteric adhesion that binds chondroitin-4-sulfate in a cooperative and shear stress–induced manner.

**Introduction**

The virulence of tropical malaria is largely associated with the bloodstream cycle of the protozoan parasite *Plasmodium falciparum* and the adhesive behavior of parasitized erythrocytes, which sequester in the deep vascular bed of inner organs to avoid passage through, and destruction in, the spleen.1 The pathological sequelae that develop in affected capillaries can progress to life-threatening complications and include diminished tissue perfusion, hypoxia, and systemic microvascular inflammation.1

Cytoadhesion of parasitized erythrocytes is mediated by a family of parasite-encoded immunvariant adhesins, collectively termed PIEmp1, that can interact with a broad range of host cell receptors, including CD36, intercellular adhesion molecule 1 (ICAM 1), P-selectin, and vascular cell adhesion molecule 1 (VCAM 1).1 A defined PIEmp1 variant encoded by the var2CSA gene can further mediate attachment of parasitized erythrocytes to chondroitin-4-sulfate (CSA). CSA is thought to be a component of the proteoglycan matrix present in the intervillous space and (moderately) on the syncytiotrophoblast cell layer of the human placenta.2-5 although one study has suggested that placental CSA is of fetal origin and is in contact with maternal blood only in the advanced and chronically infected placenta after syncytiotrophoblast denudation.6 According to this latter study, initial steps of placental cytoadhesion might involve Lewis antigens and other sialic acid–containing carbohydrates present on the intact syncytiotrophoblast and on endothelial invasive cytrophoblasts.6 Sequestration of parasitized erythrocytes in the placenta can give rise to maternal malaria, a severe syndrome associated with low birth weight, prematurity, and chronic intervillositis. PIEmp1 is presented in knoblike protrusions that anchor PIEmp1 to the erythrocyte membrane skeleton.7

Although the molecular players involved in cytoadhesion of parasitized erythrocytes are being characterized in ever-greater detail, the underpinning biomechanical principles are only partly understood. Previous studies have phenotypically described the adhesion behavior on different surfaces, revealing that parasitized erythrocytes firmly attach to CD36 and CSA but roll over surfaces containing ICAM 1, P-selectin, and VCAM 1 at physiologic wall shear stresses.8,11 Thus, central questions regarding the biomechanical factors that control the dynamics, efficiency, and strength of adhesion are still pending. The literature discusses several factors, including receptor and ligand geometry, bond characteristics, cell deformability, and the area of tight contact between the cell and the surface, that might affect the cytoadhesion of parasitized erythrocytes,12-15 but their individual or combined contribution has not yet been experimentally specified.

**Infections with the human malaria parasite *Plasmodium falciparum* during pregnancy can lead to severe complications for both mother and child, resulting from the cytoadhesion of parasitized erythrocytes in the intervillous space of the placenta. Cytoadherence is conferred by the specific interaction of the parasite-encoded adhesin VAR2CSA with chondroitin-4-sulfate (CSA) present on placental proteoglycans. CSA presented elsewhere in the microvasculature does not afford VAR2CSA-mediated cytoadhesion of parasitized erythrocytes. To address the placenta-specific binding tropism, we investigated the effect of the receptor/ligand arrangement on cytoadhesion, using artificial membranes with different CSA spacing intervals. We found that cytoadhesion is strongly dependent on the CSA distance, with half-maximal adhesion occurring at a CSA distance of 9 ± 1 nm at all hydrodynamic conditions. Moreover, binding to CSA was cooperative and shear stress induced. These findings suggest that the CSA density, together with allosteric effects in VAR2CSA, aid in discriminating between different CSA milieus. (Blood. 2015;125(2):383-391)**

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Addressing these biomechanical factors has been hampered by the lack of amenable systems. For example, it is hardly feasible to quantitatively control the receptor arrangement on a microvascular endothelial cell or on stochastically receptor-coated plastic or glass surfaces. The advent of supported membranes (planar lipid bilayers deposited on solid substrates) that can be functionalized with adhesion receptors now enables us to address these defining biomechanical principles by allowing the distance between neighboring receptors to be controlled at nanometer accuracy.\(^{16,17}\) Here, we have used functionalized membranes to study the biomechanics underpinning cytoadhesions of parasitized erythrocytes to CSA.

### Materials and methods

**P falciparum strains and cultures**

The *P falciparum* strains FCR3 and FCR3\(^{\text{var2csa}}\) have been described previously.\(^{2}\) Parasites were cultured as described and synchronized using the sorbitol method.\(^{18,19}\) FCR3 was routinely selected by gelatin flotation and repeatedly panned over CSA-coated plastic dishes or CHO-K1 cells to obtain populations adhering to CSA.\(^{20,21}\)

**Charaterization of CSA**

CSA (Sigma-Aldrich) was analyzed by 1-dimensional nuclear magnetic resonance spectroscopy. Seventy-seven percent of *N*-acetyl-D-galactosamines were sulfated at carbon C4, and 23% at carbon C6. The average molecular weight was 27.7 kDa, corresponding to a chain length of 60 disaccharide units, as determined by static light scattering (Zetasizer Nano ZS; Malvern).

**Preparation of supported membranes**

The supported membranes were prepared as described previously.\(^{16}\) Briefly, 1,2-dioleoyl-sn-glycero-3-phosphocholine (Avanti Polar Lipids) and 1,2-dioleoyl-sn-glycero-phosphoethanolamine-3-N-cap (biotinyl) (Avanti Polar Lipids) were solubilized in chloroform at molar ratios ranging from 0.02% to 5%. The lipid solutions were then dried under nitrogen flow and subsequently sonicated (0.5-second pulses for 20 minutes) with water and subsequently formed into microfluidic chambers, where a lipid bilayer was formed by vesicle fusion on the glass slides. The chambers were rinsed with water and subsequently filled with an aqueous solution containing 5 μg mL\(^{-1}\) of neutravidin (Invitrogen) to saturate the biotin tags. CSA and other glycosaminoglycans (Sigma-Aldrich) were coupled with biotin, as described previously.\(^{22}\) CSA was predominantly biotinylated at the carboxyl group of the terminal glucuronic acid, as determined by nuclear magnetic resonance. Biotinylated glycosaminoglycans at a concentration of 80 μg mL\(^{-1}\) in water were incubated with the neutravidin-saturated bilayers for 1 hour at room temperature. The functionalized membranes were subsequently washed with RPMI 1640 containing 11 mM glucose, 25 mM N-2-hydroxyethylpiperazine-N’-2-ethanesulfonic acid, and 2 mM glutamine (pH 7.6 at 37°C) (Gibco) and stored at 37°C. The intermolecular distance between CSA molecules was verified using quartz crystal microbalance with dissipation.\(^{23}\)

**Cell adhesion experiments**

Infected erythrocytes at the appropriate developmental stage were magnet purified (Milenyi Biotec), yielding parasitemias of >90%.\(^{24}\) Purified infected erythrocytes were resuspended in prewarmed complete RPMI 1640 medium at a concentration of 4 × 10\(^7\) mL\(^{-1}\), as determined using a Z1 Coulter Particle Counter (Beckman Coulter). A 180-μL cell suspension was injected into the microfluidic chamber. The filled chamber was incubated at 37°C for the time period indicated and kept in a moisture-saturated gas flow composed of 90% nitrogen, 5% oxygen, and 5% carbon dioxide during the course of the experiment. Unbound cells were washed out using a constant flow of medium at a wall shear stress of 0.8 Pa. Phase contrast images were collected using an Axiovert 200 microscope (Zeiss, Germany) equipped with an ORCA-ER charge-coupled device camera with Hokawo imaging software; Hamamatsu Photonics) mounted on a light microscope (Axiovert 200; Zeiss), original magnification ×63 (objective LD Achromat 63X/0.75 corr. Ph2; Zeiss).

**Reconstruction of height profiles**

Three-dimensional shapes of adherent cells near the membrane surface were reconstructed from 150 image stacks collected by a spinning disk brightfield confocal microscope (Nikon, Japan). The images were analyzed by the
iterative restoration method,\(^{27}\) and the reconstructed profiles were projected with z-coded stack analysis as 2-dimensional contour images.

Cell-detachment experiments

The CSA-functionalized membranes were placed in open liquid chambers, to which \(5 \times 10^6\) cells mL\(^{-1}\) were added. The chambers were incubated for 1 hour at 37°C at controlled atmospheric conditions (see above). Adherent cells were detached from the CSA-functionalized membrane by using ultrasonic pressure waves ranging from 0.3 to 5.6 MPa, with stepwise increases in increments of 0.3 MPa, generated by intensive picosecond laser pulses.\(^{28}\)

Atomic force microscopy

Atomic force microscopy on air-dried parasitized erythrocytes was performed under ambient conditions as described previously.\(^{29}\) The erythrocytes were imaged (512 \(\times\) 512 pixels) with a scanning probe microscope (NanoWizard 3; JPK Instruments), using NANOSENSORS PPP-NCLR cantilevers (force constant: 21-98 N m\(^{-1}\); resonance frequency: 146-236 kHz) with scan speeds of 0.1 to 1.0 Hz, depending on scan size (0.25-15 \(\mu\)m). All images were processed and analyzed using JPK Image Processing software.

Light scattering in solution

The size distribution of the purified VAR2CSA ectodomain (250 \(\mu\)g mL\(^{-1}\) in phosphate-buffered saline, pH 7.2) in the presence of CSA (130 and 320 \(\mu\)g mL\(^{-1}\)) or absence of CSA was determined by dynamic light scattering (Zetasizer Nano ZS; Malvern).\(^{30}\) The heterologous expression of VAR2CSA in baculovirus-infected insect cells and the purification of the protein have been described previously.\(^{31}\)

Statistics

Data were analyzed using the Student t test, analysis of variance, and the F test.

Results

CSA distance defines cytoadhesion efficiency

We initially examined the binding behavior of 3 different cell treatments on CSA-functionalized membranes (Figure 1A): (1)
erythrocytes infected with the *P. falciparum* line FCR3 preselected for high-capacity binding to CSA (henceforth called FCR3CSA); (2) erythrocytes infected with the genetically engineered clone FCR3var2CSA in which the var2csa gene exclusively conferring cytoadhesion to CSA is disrupted and which consequently does not bind to CSA; and (3) uninfected erythrocytes (adhesion to CSA-coated plastic dishes; see supplemental Figure 1 on the Blood Web site). Adhesion assays were performed using trophozoite-stage parasites (22-28 hours postinvasion). Cells were allowed to settle on the membranes (4 × 10^7 mL^-1 cells per membrane) for variable time spans (15, 30, and 60 minutes) before unattached cells were washed out.

Uninfected erythrocytes and FCR3var2CSA did not bind to the CSA-functionalized membranes, whereas FCR3CSA bound in high numbers (Figure 1B). None of the cells adhered to nonfunctionalized membranes (Figure 1B). Quantiﬁcations (Figure 1B) None of the cells adhered to nonfunctionalized membranes (Figure 1B). The number of adherent FCR3CSA further depended on the incubation time (Figure 2B), which we attribute to an increase in productive receptor/ligand interactions with time, facilitated by lateral diffusion. We fitted the data points by using a hyperbolic decay function with and without Hill coefficient. A Hill function was statistically favored in all cases, according to an F test (60 minutes: F = 16.8, P < .009; 30 minutes: F = 34.17, P < .0007; 15 minutes: F = 56.87, P < .0002). The Hill coefficients were −4 ± 1, −4 ± 1, and −6 ± 1 for the 60-, 30-, and 15-minute data sets, respectively. We further obtained values for the intermolecular CSA distance that produced half-maximal adhesion (d50). The d50 values of 9 ± 2 nm, 7 ± 2 nm, and 8 ± 2 nm for the 60-, 30-, and 15-minute data sets, respectively, were not statistically different from one another (P > .1). Neither FCR3CSA, FCR3var2CSA nor uninfected erythrocytes specifically bound to membranes functionalized with pullulan, chondroitin sulfate C, or hyaluronic acid (Figure 2C), consistent with previous studies.32

Contact area and cell deformation depend on CSA distance

The area of tight contact between parasitized erythrocytes infected with FCR3CSA and the supported membrane declined in a sigmoidal fashion with increasing CSA distance (Figure 2D). The Hill factor of −5 ± 2 and the d50 value of 6 ± 2 nm were statistically not different from the values obtained in the adhesion assays. Three-dimensional-reconstructed confocal images revealed that the magnitude of the adhesion forces deformed the bottom planes of parasitized erythrocytes (Figure 2E-F), which explains the large contact area of 9.0 ± 0.6 μm² observed on supported membranes with a CSA spacing of 6 nm.

In the case of FCR3var2CSA and uninfected erythrocytes, the contact area was independent of the CSA distance and remained constant at 2.5 ± 0.2 μm² per cell (Figure 2D). Uninfected erythrocytes maintained their biconcave-discoid shape, indicating that they did not specifically interact with the CSA-functionalized membranes (Figure 2E-F).

Assessment of detachment forces

Previous studies have estimated the adhesion strength by exposing adherent parasitized erythrocytes to gradually increasing wall shear
stresses and by determining the pressure ($\tau_{50}$) at which 50% of the cells detach. For cytoadhesion to CSA, this value is approximately 1.3 Pa. Our own wash-out experiments confirmed these studies and, in addition, showed that the CSA distance modulates the detachment process, resulting in a family of exponentially decaying curves (Figure 3A). The $\tau_{50}$ values obtained from the curves correlated in an inverse sigmoidal fashion with increasing CSA distance (Hill factor of $-11 \pm 2$ and a $d_{50}$ value of $12 \pm 2$ nm; Figure 3B).

We noted that many adherent cells were progressively deformed by the hydrodynamic pressure and eventually peeled off from the membrane. When a cell is peeled off from a surface, bonds gradually break, starting from the edge facing the pressure front. As a consequence, only relatively small pressures are required to detach a peeling cell. To obtain a more accurate estimation of the intrinsic adhesion strength, we explored an alternative detachment technique based on ultrasonic pressure waves induced by intensive picosecond laser pulses. Because of the short contact time of approximately 80 ns, cells can engage only in elastic interactions without peeling effects. Cells that were detached by this noninvasive and probe-free technique are fully viable and can readhere to surfaces. Pressures in the lower MPa range were required to detach adherent parasitized erythrocytes infected with FCR3CSA from CSA-functionalized membranes. For example, at a CSA spacing of 6 nm, 95% of the cells were detached at a critical pressure of $2.6 \pm 0.6$ MPa, as determined by analyzing the number of detachment events per pressure using a gaussian distribution (Figure 3C). The critical detachment pressure was determined for all CSA arrangements and analyzed as a function of the intermolecular CSA distance. A sigmoidal relationship emerged (Figure 3D), and fitting with a Hill function yielded a Hill factor of $-11 \pm 3$ and a $d_{50}$ value of $10 \pm 2$ nm.

In addition to trophozoites, we also investigated schizonts (32-40 hours postinvasion) to assess the effect of parasite maturation on the cytoadhesion strength. The resulting pressure diagram again revealed a sigmoidal relationship between the critical detachment pressure and the intermolecular CSA distance, but with a clear distinction. The critical pressure values were almost twice that determined for trophozoites for all CSA spacing intervals, whereas the curve parameters were largely maintained, including a Hill factor of $-11 \pm 2.0$ and a $d_{50}$ value of $12 \pm 2$ nm. Apparently, developmentally controlled parasite factors affect the cytoadhesion strength of parasitized erythrocytes, in addition to the CSA arrangement.

One parasite factor might be the knobs, which serve as focal points for PfEMP1 presentation and whose surface density increases as the parasite matures. To obtain quantitative data on knob number and distribution, we performed atomic force microscopy on erythrocytes infected with FCR3CSA. A significant increase in knob density from $9 \pm 1$ to $14 \pm 1 \mu m^{-2}$ ($P = .02$) and a decrease in basal knob diameter from $73 \pm 3$ to $64 \pm 2$ nm ($P = .004$) were observed as the parasite developed from trophozoites to schizonts (supplemental Figure 2A). The knob height remained constant ($2.8 \pm 0.3$ nm; supplemental Figure 2B). Thus, the almost twofold increase in adhesion strength as the parasite develops from a trophozoite to a schizont cannot solely be correlated with the number of knobs. Additional contributing factors might be the developmental increase in surface-presented PfEMP1, membrane-bending rigidity, or both.

Shear-induced adhesion to CSA

We investigated how a variable receptor arrangement affects cytoadhesion of parasitized erythrocytes in laminar flow. We perfused the CSA-supported membranes with cell suspensions at wall shear stresses ranging from 0.1 to 5 Pa, which covers the physiological range found in venular circulation (0.1 to 1.0 Pa, with transient peaks approaching 4.0 Pa). The total number of parasitized erythrocytes streamed through the microfluidic chambers was constant for all hydrodynamic conditions ($1 \times 10^6$). Cytoadhesion of parasitized erythrocytes was strongly dependent on the CSA distance under all hydrodynamic conditions and, in terms of the number of adherent cells, increased from none at a CSA distance of $>34$ nm to efficient adhesion at a CSA distance $<15$ nm (Figure 4A), thus replicating the findings made under static conditions. Curve-fitting each data set revealed comparable CSA $d_{50}$ values for all wall shear stress conditions, with an averaged $d_{50}$ value of $9 \pm 1$ nm (Figure 4B). Furthermore, the cooperative nature of the interaction between VAR2CSA and CSA observed under static conditions...
conditions also emerged in flow, with an average Hill coefficient of -5 ± 2. Another feature revealed by the flow experiments was the pronounced dependence of the adhesion efficiency on tensile force, in that the number of adhering cells first increased and then decreased as the wall shear stress increased (Figure 4). Optimal adhesion occurred at a wall shear stress of 0.2 ± 0.1 Pa for all CSA spacing intervals.

Antibodies against DBL domains affect cooperativity

Throughout the study, we noted cooperativity in the interaction of parasitized erythrocytes with CSA. One possibility to explain this result is that VAR2CSA can accept CSA at multiple cooperative binding sites. VAR2CSA harbors 6 DBL domains, several of which have been implicated in CSA binding, although no consensus has been reached as to whether all or only individual DBL domains interact with CSA. To investigate the putative cooperative nature of the VAR2CSA/CSA interaction, we attempted to inhibit domain-specific regions of VAR2CSA using antibodies against the individual DBL domains. We reasoned that these domain-specific antibodies might interfere with substrate-induced allosteric changes and, thus, with the degree of cooperativity. The antibodies inhibited cytoadhesion to CSA under both static and flow conditions (Figure 5A-B). Two of the 6 antibodies, namely those directed at DBL3X and DBL5, seemed to interfere with cooperative binding, indicated by significantly reduced Hill coefficients as determined in parallel assays (Figure 5C; P < .01). The CSA d₅₀ values were unaffected by the DBL domain–specific antibodies (Figure 5D).

We finally assessed the size distribution profile of the VAR2CSA ectodomain in solution. The 300-kDa protein existed in monodispersed form (Figure 6). The determined hydrodynamic radius of 9 nm was in good agreement with the calculated radius of gyration of the monomeric protein of 7 to 10 nm (depending on whether a rodlike or globular conformation was assumed). In the presence of CSA, the hydrodynamic radius shifted to a lower value of 7 nm (Figure 6), possibly because of conformational changes induced by the binding of CSA. A minor population with a hydrodynamic radius of 15 nm also emerged, likely corresponding to CSA-cross-linked ectodomains.

**Discussion**

CSA is present throughout the microvasculature, including the circulatory system of the lung and brain. Yet it is only in the placenta that the parasite *P falciparum* adheres to CSA in a process that involves specific expression of the distinct VAR2CSA adhesion molecule. This contradiction might be explained by heterogeneity in CSA populations, with only the placental intervillous space harboring the low-sulfated CSA chains of at least 6 disaccharide repeats that support adhesion of *P falciparum*–infected erythrocytes. Our data provide an additional explanation, in that parasitized erythrocytes adhere to CSA only when the average distance between neighboring sugar chains is less than 15 nm, corresponding to a minimal density of 4400 CSA molecules per square micrometer. Although the precise CSA concentration in the placental intervillous space is unknown, one can assume it to be high. CSA can be extracted in substantial quantities (>100 mg) from a single placenta, and electron microscopic studies revealed an amorphous electron-dense material in the intervillous space that was later identified as a CSA-containing proteoglycan matrix. In comparison, the CSA content on the vascular endothelium is low. CSA is predominantly attached to thrombomodulin, a transmembrane protein involved in anticoagulation. There are an estimated 50 000 to 100 000 thrombomodulin molecules on the luminal surface of an endothelial cell, of which only 10% to 20% are modified by a single CSA moiety. Assuming a luminal surface of a vascular endothelial cell between 70 and 250 μm² and a random surface distribution of the thrombomodulin molecules, this would amount to 20 to 200 CSA chains per square micrometer. These values are 1 to 2 orders of magnitude...
The ectodomain of VAR2CSA is 300 kDa in size and consists of 6 DBL domains (3 DBL domains each of the X and ε classes), a cysteine-rich interdomain region between the second and third DBL domain, and short interspersed interdomain regions. The full-length ectodomain and a subfragment containing DBL2X plus flanking sequences bind CSA with high affinity in vitro.\( ^{32,39,40} \) In addition, x-ray crystallography has suggested an interaction of CSA with the DBLX3 and the DBL6ε domains,\( ^{41-43} \) although some studies have questioned the selectivity and specificity of these DBL domains for CSA. However, these latter conclusions need to be revisited in light of our findings. If VAR2CSA is an allosteric protein, then one would expect single binding sites to have a low, if any, affinity for CSA when studied in isolation because the conformational changes through which different binding sites influence each other can take effect only in the native protein, and not in isolated binding domains. A mean Hill coefficient of 5 ± 2 suggests that most, if not all, DBL domains of VAR2CSA form binding sites for CSA. Consistent with this model, antibodies directed against DBL3X and DBL5ε significantly reduced the degree of cooperativity.

Adhesion of parasitized erythrocytes to CSA displays an interesting phenotype under variable hydrodynamic conditions, in that the amount of adhering cells first increases and then decreases as the wall shear stress is increased. Shear-enhanced adhesion is well-recognized in other systems, including FimH-mediated adhesion of Escherichia coli bacteria and selectin-mediated adhesion of leukocytes, and is explained by catch bonds—receptor/ligand interactions that are strengthened by tensile force owing to force-induced allosteric effects that prolong the half-life of the interaction.\(^{50,51} \) On the basis of our hydrodynamic experiments, we propose that the interaction between VAR2CSA and CSA behaves like a catch bond when stressed. Shear-enhanced adhesion was also noted for rolling adhesion of parasitized erythrocytes to ICAM-1,\(^8,10 \) suggesting that it might be a feature common to different PfEMP1/receptor interactions. The wall shear stresses of 0.2 to 0.8 Pa that afforded adhesion to CSA correspond well with hydrodynamic conditions allegedly present in the intervillous space.\(^{52,53} \)

Wash-out assays have been widely used to assess the adhesion strength of parasitized erythrocytes to CSA and other host receptors.\(^8,11,33 \) Although such assays mimic physiological hydrodynamic conditions, they grossly underestimate the intrinsic strength of the cell/receptor interaction. When adherent cells are exposed to increasing wall shear stresses, they deform under the hydrodynamic pressure and eventually peel off as bond after bond breaks. Removing adherent cells from a surface by using a micropipette or the cantilever of an atomic force microscope results in similar effects.\(^8,13,54 \) In comparison, ultrasonic pressure waves do not permit peeling effects or cell deformation. Because the noncorpuscular waves propagate at an ultrasonic speed of approximately 1640 m s\(^{-1} \), the contact time is extremely short (approximately 80 ns), thereby limiting the effect on the cell to elastic interactions.\(^{54} \) Thus, for the cell to detach from the contacting surface, all bonds need to rupture instantaneously and simultaneously. Consequently, pressures much higher than those used in wash-out assays are needed to detach cells. Further, the high loading rates applied in the laser ablation assay result in stronger interactions between noncovalent ligand/receptor pairs and, hence, higher unbinding forces.\(^{55} \)

The detachment of parasitized erythrocytes from CSA is highly cooperative in a manner reminiscent of subunit cooperativity as is observed, for example, when a DNA helix melts. This finding suggests that the periodic pattern in which the CSA molecules are

![Figure 6. Size distribution profile of the VAR2CSA ectodomain in the presence and absence of CSA in solution.](image-url)
presented on the supported membrane is matched by a similar periodic organization of the VAR2CSA molecules, suggesting that VAR2CSA, and possibly other PIEMP1 variants, are presented on the knobs in a defined geometric arrangement. In summary, our data suggest that the intermolecular CSA distance, the cooperative nature of VAR2CSA/CSA interaction, and the shear-enhanced binding behavior are defining parameters for the specific and selective cytoadhesion of \textit{P} \textit{falciparum}–infected erythrocytes in the placental intervillous space.

Acknowledgments

The authors thank Stefan Prior for technical assistance. M.T. and M.L. are members of the German Excellence Cluster “CellNetwork.” M.T. and M.L. are further members of the Helmholtz Program “BioInterface” and the European Network of Excellence “EvMalR,” respectively.

This work was supported by the Deutsche Forschungsgemeinschaft under the Sonderforschungsbereich 1129 and the European Seventh Framework Program grant “PreMalStruct.” The Institute for Integrated Cell-Material Science is supported by the World Premier International Research Center Initiative, Ministry of Education, Culture, Sports, Science, and Technology, Japan. M.T. and H.Y.Y. thank the Japan Society for the Promotion of Science for the support by Kakenhi (grants 26247070 and 24680050) and the Brain Circulation Program, respectively.

Authorship

Contribution: M.L. and M.T. conceived the study, analyzed the data, and wrote the manuscript; H.R., H.Y.Y., K.Q., M.A.N., C.P.S., and A.S. performed the experiments and analyzed the data.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

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References


high-affinity binding to CSA. Proc Natl Acad Sci USA. 2010;107(11):4884-4889.


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